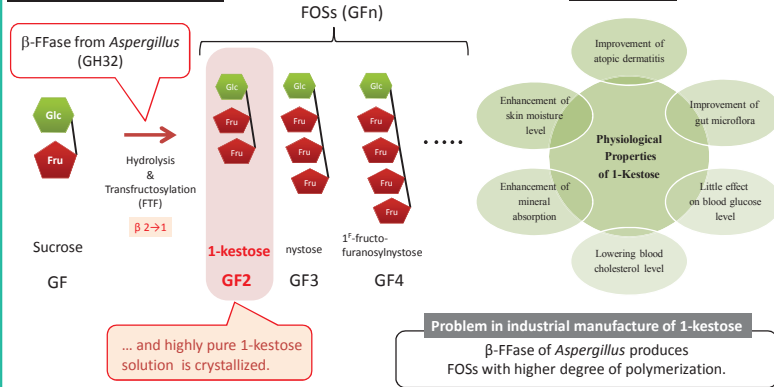


## 1. Abstract

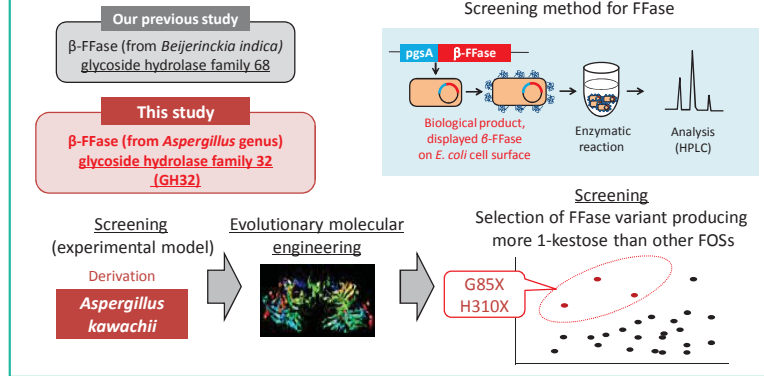
1-Kestose is a component of **fructooligosaccharides (FOSs)** and a food ingredient with a significant commercial value. 1-Kestose is produced by  **$\beta$ -fructofuranosidase (FFase)**, a **transfructosylating (FTF)** and hydrolytic enzyme.  $\beta$ -FFases of *Aspergillus* are generally used for the production of FOSs. However, these enzymes are not suitable for 1-kestose production because it undergoes rapid polymerization to FOSs. In this study, we adapted  $\beta$ -FFase derived from *Aspergillus kawachii*, belonging to **glycoside hydrolase family 32 (GH32)**, as an experimental model and engineered this  $\beta$ -FFase for efficient production of 1-kestose using evolutionary molecular engineering techniques. Our study will lead to construction of an industrial enzyme producing 1-kestose efficiently.

## 2. Introduction

### What is 1-kestose ?



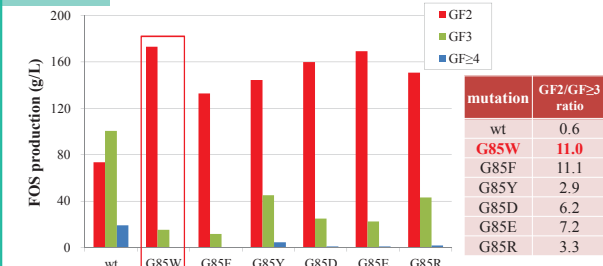
## 3. Outline of the study



## 4. Purpose

To engineer  $\beta$ -FFase producing 1-kestose efficiently.

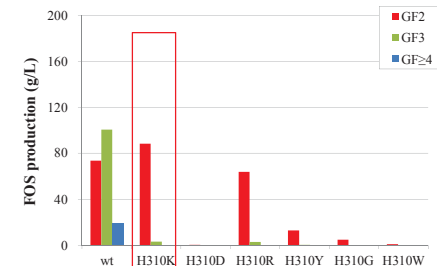
## 5. Results



**Fig. 1 Evaluation of FOS production in *A. kawachii* FFase G85X mutants**

[Sucrose conc. 30% [w/w], 0.35 mL, 30°C, pH7, 3 h]

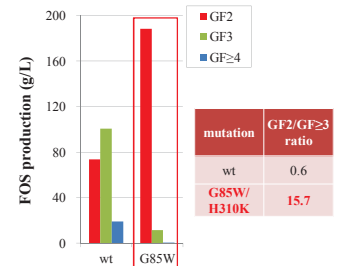
- The production of GF2 and GF $\geq$ 3 was higher and lower in the G85X mutants than in the wild-type, respectively.
- G85W mutants effectively produced more GF2 and less GF $\geq$ 3.



**Fig. 2 Evaluation of FOS production in *A. kawachii* FFase H310X mutants**

[Sucrose conc. 30% [w/w], 0.35 mL, 30°C, pH7, 3 h]

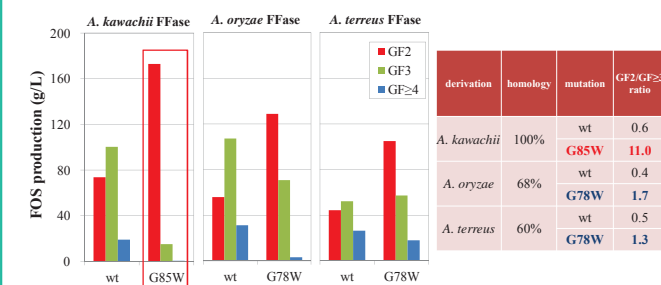
- The production of GF2 and GF $\geq$ 3 was higher and lower in H310K mutant than in the wild-type, respectively.
- GF $\geq$ 3 was hardly produced in all H310X mutants.



**Fig. 3 Evaluation of FOS production in *A. kawachii* FFase G85W/H310K mutant**

[Sucrose conc. 30% [w/w], 0.35 mL, 30°C, pH7, 32 h]

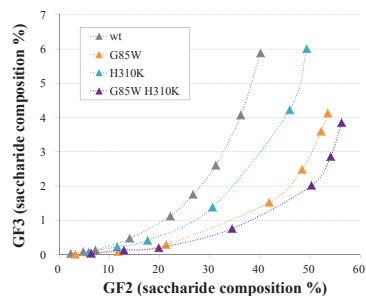
- The production of GF2 and GF $\geq$ 3 was higher and lower in G85W/H310K mutant than in the wild-type and single mutants, respectively.



**Fig. 4 Evaluation of FOS production in homologs of *A. kawachii* FFase, belonging to GH32**

[Sucrose conc. 30% [w/w], 0.35 mL, 30°C, pH7, 3 h]

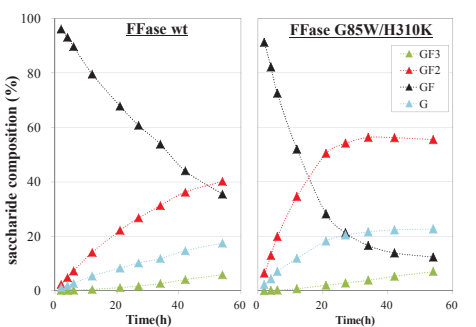
- According to sequence alignment of  $\beta$ -FFases, belonging to GH32, G85 of *A. kawachii* FFase was conserved as G78 of *A. oryzae* and *A. terreus* FFases, respectively.
- In *A. oryzae* and *A. terreus* FFase G78W mutants, the production of GF2 and GF $\geq$ 3 was not very different from that in the wild-type, respectively.



**Fig. 5 GF2 and GF3 production profiles of *A. kawachii* FFase mutants**

[Sucrose conc. 30% [w/w], 0.4 mL, 30°C, pH7, over time]

- According to the GF2 and GF3 production profiles in the saccharide composition, GF2 and GF3 were more and less in all mutants, especially the double mutant, than in the wild-type, respectively.

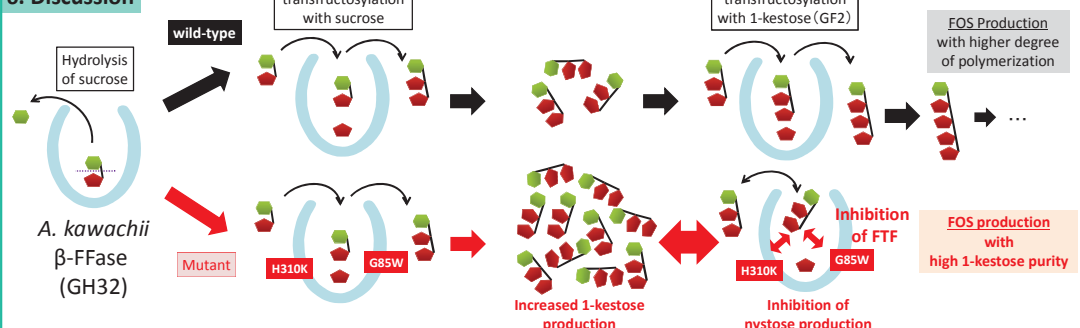


**Fig. 6 Saccharide composition in *A. kawachii* FFase wt and G85W/H310K over the course of time**

[Sucrose conc. 30% [w/w], 0.4 mL, 30°C, pH7, over time]

- The specific FTF activity from sucrose to GF2 in double mutant was higher than that in the wild-type.

## 6. Discussion



## 7. Conclusion

We concluded that G85 and H310 of *A. kawachii* FFase were involved in the regulation of FTF from 1-kestose to nystose. Their FFases may enable efficient 1-kestose production.

## 8. Future perspectives

Our study will enable development of an industrial enzyme producing 1-kestose efficiently, which is generally considered difficult.